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The Synthesis of 7-Carbonyl Homologues of 1-Deoxynojirimycin

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Abstract: A 3,4;5,6-di-*O*-isopropylidene salt derivative of 1-amino-1-deoxy-D-glucitol was transformed to *N*-Boc protected (5,6)- α,β -unsaturated 7-carbonyl compounds. Conversion to the title piperidine products proceeded *via* deprotection and intramolecular 1,4-addition of the amino group.

Inhibition of glycosidases may be useful for treatment of several diseases e.g. diabetes,¹ cancer,² and some viral infections.³ Important examples are the antiviral (including anti-HIV) and antidiabetic activities⁴ found for the glucosidase inhibitors 1-deoxynojirimycin⁵ **1** and castanospermine⁶ **2** (Figure 1). The potential chemotherapeutic applications of these natural polyhydroxylated alkaloids and their analogues have prompted considerable synthetic interest towards structural modification, such as the introduction of lipophilic (e.g. fluoro,⁷ alkyl,⁸ and acyl⁹), amino,^{10,11} and glucosyl¹² groups at specific positions of compound **1**. Complete removal of the C-6 hydroxymethyl group of **1** has remarkably little effect on enzyme-substrate interactions.¹³

This report deals with the conversion of 1-amino-1-deoxy-D-glucitol **3** to the trihydroxypiperidines **4-7**. These compounds represent a new class of 1-deoxynojirimycin analogues where the C-6 hydroxyl has been replaced with a ketone, acid or amide carbonyl function.

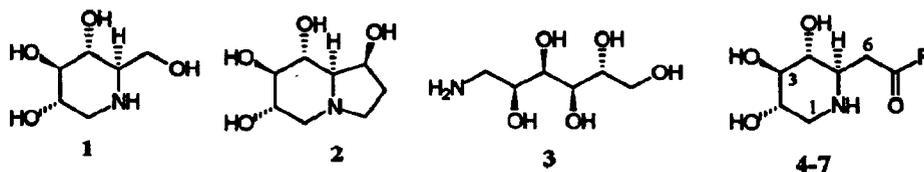
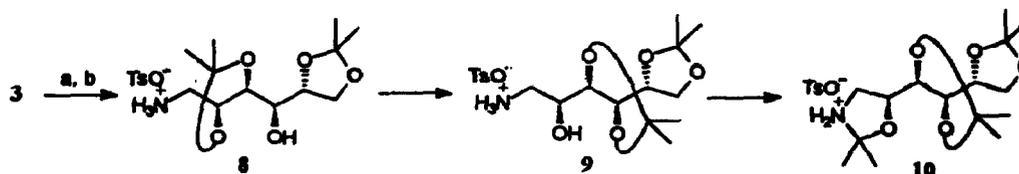


Figure 1

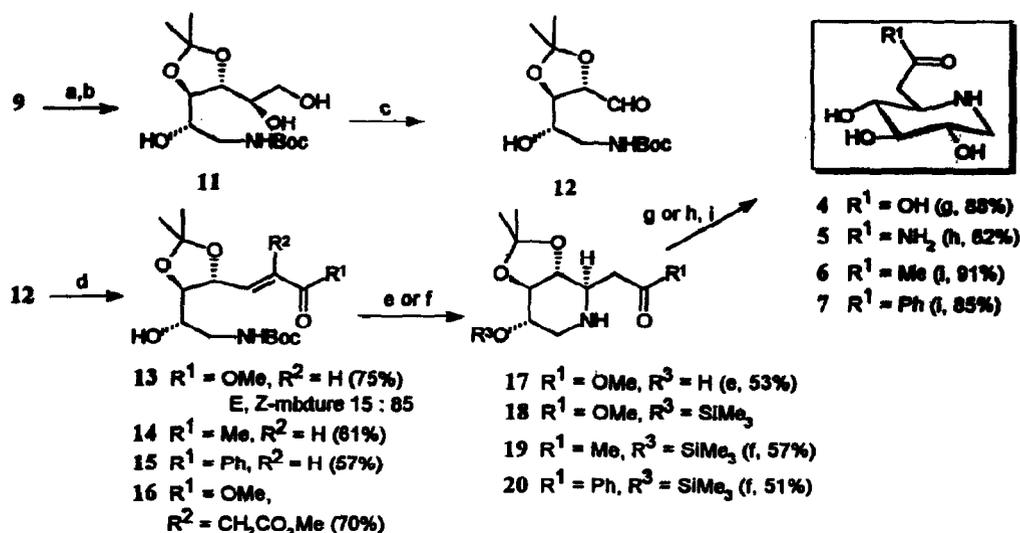
The synthesis started with the preparation of the 3,4;5,6-di-*O*-isopropylidene protected salt **9**. The course of the reaction depicted in Scheme 1 was revealed by t.l.c analysis of the *N*-acrylynyl [RNHCH=C(CO₂Et)₂] and *N*-Boc derivatives.¹⁴ Under the acidic conditions, the initially formed diacetonide **8** rearranged to the regioisomer **9** which crystallized from the reaction medium. After 24 hours, pure compound **9** was collected by filtration (65% yield), whereas in the filtrate only diacetonide **9** and triacetonide **10** were detected.

Scheme 1.



Experimental conditions: (a) MeOH, *p*-TsOH.H₂O (1 eq), evaporate to dryness; (b) Me₂CO-Me₂C(OMe)₂ (1:3), *p*-TsOH.H₂O (0.5 eq), r.t. 24 h, 65% **9**.

Scheme 2



Experimental conditions: (a) PPTS (1 eq), MeOH-H₂O (9:1), 60 °C, 1 h; (b) Na₂CO₃, (*t*-BuOCO)₂O, 0.5 h; (c) NaIO₄ (1.1 eq), H₂O; (d) R¹COCR²=PPh₃, (1.2 eq), MeOH or CH₂Cl₂, 0.5 h; silica column (EtOAc-hexane 1:1); (e) Me₃SiI, CH₂Cl₂, r.t., 10 min; MeOH-Et₃N; MeOH-K₂CO₃; silica column (EtOAc-MeOH 94:6); (f) Me₃SiI, CH₂Cl₂, r.t., 5 min; Et₃N; silica column (EtOAc-hexane 3:2); (g) 6M HCl; Dowex 50W-X8 (0.2M NH₄OH); (h) HCl-MeOH; NH₃-MeOH, NaCN, reflux; Dowex 50W-X8 (0.2M NH₄OH); (i) 6M HCl; silica column (CHCl₃-MeOH-H₂O-NH₄OH 70:28:1:1).

Diacetonide **9** was converted (Scheme 2) to the crystalline *N*-Boc protected 2,5,6-triol **11** in 81% yield.¹⁵ Selective removal of the 5,6-*O*-isopropylidene group was achieved by heating **9** with pyridinium *p*-toluenesulfonate (PPTS) in aqueous methanol at 60 °C. After reaction with (*t*-BuOCO)₂O, triol **11** was separated from the *N*-Boc derivative of the remaining diacetone **9** by successive extraction with toluene and ethyl acetate. The oxidative cleavage of the 5,6-diol group of triol **11** with NaIO₄ afforded the unstable L-xylose derivative **12**, which was directly subjected to various Wittig reactions in methanol or dichloromethane. By using the appropriate triphenylphosphoranylidene reagents,¹⁶ the α,β -unsaturated carbonyl compounds **13-16** were isolated in 57-75% yield based on the triol precursor **11**.¹⁷

Our synthetic plan (Scheme 2) required deprotection of the *N*-Boc amino group and cyclization via 1,4-addition to the α,β -unsaturated carbonyl function. Treatment of the ester compound 13 with formic acid for 10 minutes and neutralisation with aqueous Na_2CO_3 gave the desired piperidine compound 17 in only 35% yield. The low yield was due to incomplete deprotection of the amino group and, presumably, to partial cleavage of the isopropylidene group resulting in detection of unidentified polar side products. A more selective deprotection was accomplished by reaction with Me_3SiH in dichloromethane and quenching with methanol and Et_3N . Under these conditions, a mixture of the cyclic compound 18 and the desilylated product 17 was isolated. No cyclization was observed in the absence of methanol. Complete removal of the trimethylsilyl group with K_2CO_3 in methanol produced 17 in 53% overall yield from 13.

In the analogous conversion of ketones 14 and 15 with Me_3SiH , cyclization of the intermediate primary amines already occurred upon addition of only Et_3N to the reaction mixture. However, for diester compound 16 none of the expected monocyclic and/or bicyclic product was observed on removal of the *N*-Boc group and heating of the resulting primary amine in methanol or 2-butanol.

Conversion of compounds 17-20 to the target compounds 4-7 proceeded via acidic cleavage of the protecting groups. Treatment of ester 17 with aqueous 6M HCl for 48 hours followed by ion exchange chromatography provided the crystalline amino acid 4. The amide 5 was prepared from 17 by sequential removal of the acetal group with methanolic HCl and ammonolysis of the resulting ester intermediate. The ammonolysis was effected by prolonged heating with methanolic ammonia using NaCN as a catalyst.¹⁸ After acidic deprotection of 19 and 20 with 6M HCl, the ketone target compounds 6 and 7 were isolated by column chromatography on silica gel.¹⁷

Analysis of the ^1H NMR spectra of the *trans*-fused acetonides 17-20 and the deprotected compounds 4-7 revealed the all-equatorial orientation of the substituents, as shown by the coupling constant values $J_{5,4} = J_{4,3} = J_{3,2} = 9$ Hz.¹⁹ Hence, 1,4-addition of the amino group to the α,β -unsaturated carbonyl system proceeds in a diastereospecific way producing the C-6 equatorial isomer exclusively.

The acetonides 17-20 represent advanced intermediates showing the desired variety of protected and non protected functional groups. When further modified and deprotected, they may provide access to a large number of variously substituted analogues, in addition to the C-6 extended homologues 4-7 resulting from simple hydrolysis.

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 14. Samples of the reaction mixture were treated with aqueous Na₂CO₃ and diethyl ethoxymethylenemalonate or di-*tert*-butyl dicarbonate. T.l.c. analysis (hexane-EtOAc, 7:3) revealed the *N*-acylvinyl derivatives of **8** and **9** (**8** more polar than **9**) or the corresponding *N*-Boc derivatives (R_F **8** = R_F **9** = 0.2) and the *N*-Boc triacetone **10** (R_F = 0.5). The secondary amine does not react with the acylvinyl reagent.
 15. Crystallized from hexane-EtOAc, mp 93-94 °C, [α]_D¹⁸ +8.57° (c 0.2, MeOH). HRMS for tris-*O*-trimethylsilyl derivative, calcd. for C₂₂H₄₈NO₇Si₃ ([M-CH₃]⁺) 522.2739, found 522.2741. The 81% yield for triol **11** includes material (28%) obtained by subjecting the *N*-Boc diacetone fraction resulting from non hydrolyzed **9** to the PPTS acid hydrolysis.
 16. The reactions were conducted in MeOH except for the use of CH₂Cl₂ in the preparation of **16**. The commercial reagents RCOCH=PPh₃ (R = OMe, Me) were used. PhCOCH=PPh₃ was generated *in situ* by treatment of commercial phenacetyltriphenylphosphonium bromide with MeONa. The reagent MeO₂C-C(CH₂CO₂Me)=PPh₃ was prepared by using a modification of the method reported by Cameron, A. F.; Duncanson, F. D.; Freer, A. A.; Armstrong, V. W.; and Ramage, R.; *J. Chem. Soc. Perkin Trans II* **1975**, 1030.
 17. Satisfactory spectral data (IR, 400 MHz ¹H NMR, ¹³C NMR, EI and CI mass spectra) were obtained for all new compounds. HRMS data were acquired for all new compounds (M⁺ or significant fragment ions were measured). Compounds **4**, **6**, and **11** were analysed as the trimethylsilyl derivatives.
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 19. Selected NMR data : **4** ¹H NMR 400 MHz (D₂O), δ (ppm) 2.11 (dd, J = 9, 16 Hz, 1 H, H-6a), 2.47 (dd, J = 11, 12 Hz, 1 H, H-1ax), 2.72 (dd, J = 3, 16 Hz, 1 H, H-6b), 2.73 (m, 1 H, H-5), 3.06 (dd, J = 5, 12 Hz, 1 H, H-1eq), 3.08 (t, J = 9 Hz, 1 H, H-4), 3.28 (t, J = 9 Hz, 1 H, H-3), 3.48 (ddd, J = 5, 9, 12 Hz, 1 H, H-2); ¹³C NMR 100 MHz 39.7 (C-6), 48.9 (C-1), 57.5 (C-5), 70.6 (C-2), 74.4 (C-4), 78.3 (C-3), 180.2 (COOH). **5** ¹H NMR 400 MHz (D₂O), δ (ppm) 2.18 (dd, J = 9, 16 Hz, 1 H, H-6a), 2.40 (dd, J = 11, 12 Hz, 1 H, H-1ax), 2.71 (dd, J = 3, 16 Hz, 1 H, H-6b), 2.78 (td, J = 3, 9 Hz, 1 H, H-5), 3.02 (dd, J = 5, 12 Hz, 1 H, H-1eq), 3.05 (t, J = 9 Hz, 1 H, H-4), 3.25 (t, J = 9 Hz, 1 H, H-3), 3.43 (ddd, J = 5, 9, 12 Hz, 1 H, H-2); ¹³C NMR 100 MHz 37.3 (C-6), 48.6 (C-1), 56.8 (C-5), 70.5 (C-2), 74.2 (C-4), 77.8 (C-3), 176.6 (CONH₂). **6** ¹H NMR 400 MHz (D₂O), δ (ppm) 2.15 (m, 1 H, CHDCO), 2.17 (s, 1 H, CD₂HCO), 2.42 (t, J = 11, 12 Hz, 1 H, H-1ax), 2.87 (m, 1 H, H-5), 3.01 (dd, J = 5, 12 Hz, 1 H, H-1eq), 3.07 (t, J = 9 Hz, 1 H, H-4), 3.24 (t, J = 9 Hz, 1 H, H-3), 3.44 (ddd, J = 5, 9, 12 Hz, 1 H, H-2); ¹³C NMR 100 MHz 29.8 (CH₃CO), 44.5 (CH₂CO), 48.5 (C-1), 55.7 (C-5), 70.3 (C-2), 73.9 (C-4), 77.8 (C-3), 213.4 (CO). **7** ¹H NMR 400 MHz (CD₃OD), δ (ppm) 2.60 (br t, J = 12 Hz, 1 H, H-1ax), 3.13 (m, 1 H, H-5), 3.16 (dd, J = 5, 12 Hz, 1H, H-1eq), 3.22-3.33 (m, 2 H, H-3, H-4), 3.56 (m, 1 H, H-2); 7.50, 7.62, 8.01 (m, 5 H, Ph); ¹³C NMR 100 MHz 40.4 (C-6), 50.7 (C-1), 58.0 (C-5), 71.7 (C-2), 75.0 (C-4), 80.1 (C-3), 129.3, 129.8, 134.7, 138.1 (C arom.), 201.0 (CO).